



Electrospun Membrane Fabrication in Assisting Tissues Healing

Madeeha Sadia^{1,2}, Norhidayu Muhamad Zain³, Nurizzati Mohd Daud¹, Faizuan Abdullah⁴, Ali Dad Chandio⁵, Iftikhar Ahmed Channa⁵, Syed Ali Ammar Taqvi⁵, Nik Ahmad Nizam Nik Malik⁴, Syafiqah Saidin^{1,6,*}

¹Department of Biomedical Engineering & Health Sciences, Faculty of Electrical Engineering, Universiti Teknologi Malaysia, 81310 UTM Johor Bahru, Johor, Malaysia.

²Department of Biomedical Engineering, Faculty of Electrical and Computer Engineering, NED University of Engineering and Technology Karachi, Pakistan.

³Academy of Islamic Civilization, Faculty of Social Sciences and Humanities, Universiti Teknologi Malaysia, 81310 UTM Johor Bahru, Johor, Malaysia.

⁴Department of Chemistry, Faculty of Science, Universiti Teknologi Malaysia, 81310 UTM Johor Bahru, Johor, Malaysia.

⁵Faculty of Chemical & Process Engineering, NED University of Engineering and Technology Karachi, Pakistan.

⁶IJN-UTM Cardiovascular Engineering Centre, Institute of Human Centered Engineering, Universiti Teknologi Malaysia, 81310 UTM Johor Bahru, Johor, Malaysia.

*Corresponding Author syafiqahs@utm.my

Received 06 June 2023; Accepted 17 July 2023; Available online 06 August 2023

<https://doi.org/humentech.v2n2.55>

Abstract:

The arrangement of fibers into three-dimensional (3D) complex structure is constructing a sheet of membrane, depending on the fabrication technique. The fiber mats and scaffolds have been used in various applications including tissue engineering which involve the integration of cells and tissues within the pores between the fibers. There are several techniques that have been opted to produce specifically nanofibers as its efficacy in tissue healing is prominent compared to microfibers. Among the fabrication techniques (drawing, template synthesis, temperature-induced phase separation, molecular self-assembly, and electrospinning), electrospinning method has drawn attention due to their easy handling, inexpensive, and ability for membrane scale-up with the production of fibers ranging from few nanometers to several microns. Researchers have employed a variety of electrospinning methods, including blended/co-electrospinning, emulsion, coaxial, side-by-side, and triaxial electrospinning. In electrospinning smooth formation of nanofibers for tissue healing with less appearance of spray and beads, several parameters such as humidity, temperature, voltage, flow rate, viscosity, concentration, molecular weight, surface tension, conductivity, and solvent volatile need to be tailored. The morphology of nanofibers formation should support the size and structure of the surrounded cells and tissues. Besides, the types of degradable polymeric materials also play a role in the formation of stable nanofibers. This review paper aimed to provide information on the techniques to produce nanofibers, intended to the basic exploration of electrospinning.

Keywords: Electrospinning; Membrane; Nanofibers; Tissue healing

1. Introduction

Nanofibers are among the most intriguing and promising materials due to their special physicochemical characteristics, such as remarkable porosity with interconnection between pores in mats, mechanical flexibility and strength, large surface area, and high adaptability to construct composites with other materials [1, 2]. These features permitting its usage for a range of applications have generated enormous scientific and technical attention [1, 2]. The creation of nanofibers has thus far involved techniques including drawing [3], self-assembly [4], phase separation [5], and template synthesis [6]. They are time-consuming, expensive, and inefficient, which are some of their drawbacks. However, electrospinning-based designed electrospun membranes/nanofibers/scaffolds have distinctive features which make them quite appropriate for drug delivery system [7], tissue engineering scaffolds/membranes, and biomedical engineering [8].

Table 1 shows the comparison of advantages and disadvantages of different nanofiber techniques which includes drawing, template synthesis, temperature induced phase separation, molecular self-assembly [7, 9, 10]. These techniques have contained some main drawbacks which are time-consuming, expensive, and inefficient. Polymeric fibrous structures are identified as suitable candidates for drug-release systems and fibrous polymer structures. They have been prepared by a variety of methods such as electro-hydrodynamic (EHD) techniques [11]. In the EHD, electrostatic forces are employed to produce fibers or particles with an adjustable micro-structure [12]. As an outstanding cost-efficient method, electrospinning is categorized as one of the EHDs, that has been opted in vast applications in the industry and laboratory for fiber fabrication. In this method, pharmaceutical biomolecules or agents are directly encapsulated within the fibers, leading to their protection from environmental parameters and simultaneously, controlling the element release [13, 14]. Electrospinning is the famous technique which followed the working principle of EHD [11]. Electrospinning-based designed electrospun membranes/nanofibers/scaffolds have distinctive features which make them quite appropriate for drug delivery system, tissue engineering scaffolds/membranes and biomedical engineering. Electrospinning technique have covers unique characteristics which includes high porosity, tuneable pore size, high surface-to-volume ratio, and morphological correspondence with extracellular matrix (ECM) [10].

Table 1 - Comparison of different nanofiber techniques [7, 9].

Fabrication Technique	Advantages	Disadvantages
Drawing	Easy-to-handle equipment	Inconsistent process Not expandable Fiber diameters are not under the control
Template synthesis	Continuous process Fiber dimension can be varied using different template	Not scalable
Temperature-induced phase separation	Easy-to-handle equipment Easy processing Formation of fibers with good mechanical characteristics	Fabrication only involves polymer materials No scalable Unable to control fiber dimension
Molecular self-assembly	Only smaller nanofibers of few nm in diameter and few microns in length can be fabricated	Complicated process No scalable Unable to control fiber dimension
Electrospinning	Easy instrument handling Continual procedure Inexpensive Scalable Fabrication of fibers with sizes ranging from few nanometers to several microns	Instability in jet formation Certain harmful solvents Transportation, handling, and packaging

With the help of synthetic and natural polymers, it is simple to create antibacterial drug-loaded [7] and wound-healing nanofibers [15, 16] that are then electrospun [10]. Numerous antibacterial substances, including antibiotics, metallic nanoparticles, and proteins have been fabricated, to be contained within or on the surface of nanofibers [10, 17]. Previous research have demonstrated the potent antibacterial activity of electrospun membranes fabricated with natural and synthetic polymers such as gelatin [18], cellulose [19], chitosan [20], collagen [21], poly (methyl methacrylate) [10], polyvinylpyrrolidone (PVP) [22, 23], and polycaprolactone (PCL) [24] against a wide range of bacteria with strong biocompatibility for wound healing applications [10]. Therefore, this review paper aimed to

provide information on the techniques to produce nanofibers, intended to the basic exploration of electrospinning. The types of electrospinning specified parameters and dominant polymeric materials have been reviewed to provide an insight on its application towards tissue healing.

2. Electrospinning

Researchers have employed a variety of electrospinning methods, including blended/co-electrospinning (single fluid), emulsion electrospinning (single fluid), coaxial electrospinning (double fluid), side-by-side electrospinning (double fluid), and triaxial electrospinning (multi-fluid) as shown in Figure 1. Particularly to load-sensitive molecules or small water-soluble drugs, these strategies aid in controlling drug release from nanofibers, and support high surface area with improved morphological and biological properties in the required environment [10].

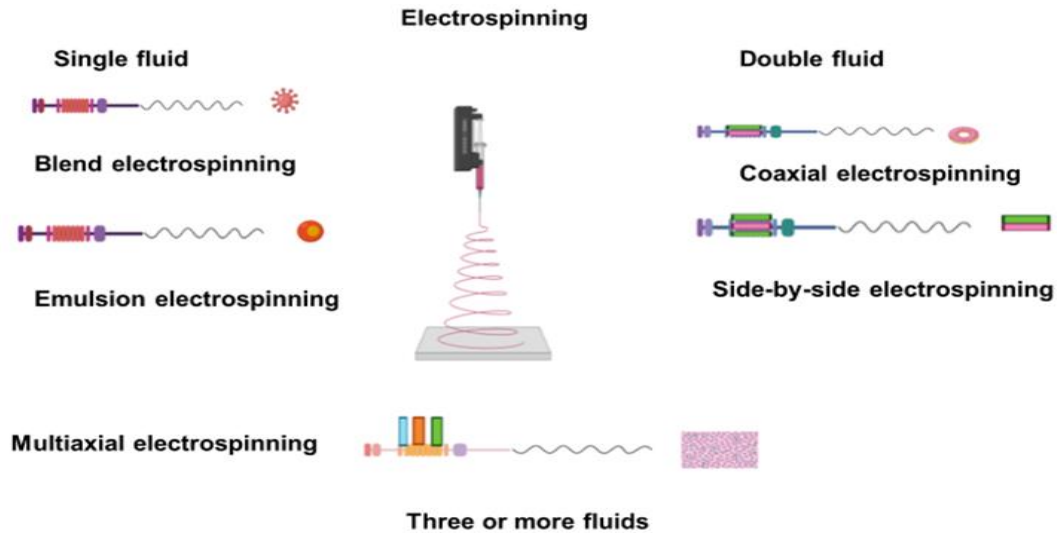


Figure 1 Illustration of different types of electrospinning methods

Co-electrospinning is a flexible and easy method for creating nanofibers of various polymers with the capacity to load a variety of molecules in these structures [25, 26]. Emulsion electrospinning is an easy and common technique for creating core-shell nanofibers, which are good for loading drugs [27]. The hydrophilic pharmaceuticals in emulsion electrospinning are typically dissolved in the water phase before being diffused into the oil phase, which contains surfactants/emulsifiers, in contrast to the blend electrospinning discussed above [28]. After electrospinning, the water/oil emulsion creates nanofibers with a core-shell structure, the core of which contains the drug [29]. As a result, it exhibits higher performance in the ability to encapsulate physiologically active molecules, such as proteins and medications [25].

The emulsion technique does not need a specific needle setup, but it does need a certain combination of solvent, polymers, and molecular characteristics. In the coaxial/multiaxial approach, the initial bioactivity of molecules is maintained by electrospinning without direct mixing the solution by utilizing core-shell nozzles [30]. The method of side-by-side electrospinning for creating Janus nanofibers was initially described by Gupta and Wilkes in 2003 [31]. The Janus structure is among the most fundamental ones. The two chambers of this construction are distinct from one another and in contact with the outside world, unlike the conventional core-sheath configuration. By figuring out the spinneret's structure and modifying the electrospinning settings, side-by-side electrospinning allows the fabrication of nanofibers with various characteristics [32].

By using a multi-layered nanofiber, each of which can perform a separate function while maintaining high mechanical stability. This double-layered approach can offer multifunctionality [33]. Hence in this context, it is concluded that coaxial emulsion, side-by-side, and multiaxial procedures are more difficult to be handled than blended electrospinning, and careful consideration of material choice, solvent combination, and particular volatility are required [10, 25]. Blended/co-electrospinning was adopted in this study to create polymer-based loaded leave extract and metallic nanoparticles electrospun membranes as it is a versatile and simple method.

Scientists have loaded plant extracts and bioactive ingredients within PCL by opting blended/co-electrospinning technique to produce various electrospun membranes with improved properties for wound healing applications. The recently reported works were PCL/chitosan/Aloe vera [34], PCL/chitosan/curcumin [35], PCL/gelatin/oregano oil [36], PCL/gelatin/clove essential oil [37], and PCL/gelatin/Utrica dioica/zinc oxide nanoparticles [38]. All membranes exhibited improved antibacterial and biocompatible properties for wound healing applications.

2.1 Electrospinning parameters

The structure and development of fibers produced by the simple technique of electrospinning are influenced by several factors. According to Table 2, the variables are broken down into three groups: ambient parameters, processing parameters, and solution parameters. All three groups should be adjusted to create smooth electrospun fibers.

Table 2 - Important parameters used during electrospinning.

Parameter	Effect	Parameter Value	Reference
Ambient Parameters			
Humidity	<ul style="list-style-type: none"> High: Fiber diameter and pore size increased. Low: Beads formation. Smooth: Fibers formed 20 to 75%. 	Between 50 - 60%	[39-42]
Temperature	<ul style="list-style-type: none"> High: The rate of liquid evaporation increased, viscosity decreased, increase in temperature encouraged the formation of thin fibers. Low: Chances of fiber creation trapped at the needle end 	Room temperature (25 - 27°C)	[42-45]
Processing Parameters			
Voltage	<ul style="list-style-type: none"> High: Facilitated formation of large fiber diameter. Low: Thin diameter. Mixed: Affected fiber morphology due to polymer concentration, flow rate, the distance between needle and collector with applied voltage. 	10 kV	[42, 46-48]
Flow rate	<ul style="list-style-type: none"> Low: Preferable for polymer polarization and thinner fiber formation. High: Beads formation occurred, and fiber diameter became thicker. 	1 mL/h	[40, 42, 49]
Solution Parameters			
Viscosity	<ul style="list-style-type: none"> Depended on polymer concentration and molecular weight. Low: No fiber formation, electrospaying Slightly low: Formed beads and depended on surface tension. Very high: Led to the hard solution being pushed out from the jet due to high surface tension. High: Produced large fiber diameter. 	15 wt% polymer solution	[41, 42, 48, 50]
Concentration	<ul style="list-style-type: none"> Adjustable concentration is required to facilitate smooth fibers formation. Low: No fiber formed. High: Beads formation 	Different polymer solution concentrations	[42]
Molecular weight	<ul style="list-style-type: none"> Supported the entanglement of polymer chains in solution. Low: Facilitated beads formation. High: Produced smooth electrospun membranes 	80 kDa	[41, 42, 47, 49]
Surface tension	<ul style="list-style-type: none"> High: Inhibited electrospinning Low: Allow electrospinning process, depended on the selected solvent and modification of the 	-	[42, 46, 48, 50]

Parameter	Effect	Parameter Value	Reference
	ratios of solvents surface tension as well as viscosity		
Conductivity	<ul style="list-style-type: none"> • High conductivity is required to overcome surface tension and to produce bead less fibers. • Poor conductivity produced beads without the formation of fibers. Conductivity can be increased by adding salts, and drugs. 	-	[41, 42, 47, 49, 51]
Solvent volatility	<ul style="list-style-type: none"> • A volatile solvent is recommended. • Prevent to use of extremely volatile solvents due to the fast evaporation rate and the solution may clog the needle. • Solvent evaporation is needed before reach to the collector otherwise it may merge and produce beads 	Chloroform Methanol	[41, 51, 52]

2.2 Electrospinning Materials

Over natural polymers (biopolymers), synthetic polymers have several benefits, with the most significant benefit to modify the materials' chemical characteristics, molecular weight, copolymerization, crystallinity, and other characteristics to achieve better mechanical and degrading qualities [53, 54]. For wound dressing formulations, synthetic polymers that can be electrospun with natural polymers include poly(vinyl alcohol) (PVA), PLGA, polylactide (PLA), PCL, polyglycolic acid (PGA), polyurethane (PU), poly(ethylene oxide) (PEO)/poly(ethylene glycol) (PEG), poly(hydroxyethyl methacrylate) (PHEMA), and poly(vinyl pyrrolidone) (PVP) [52, 53, 55].

Additionally, the cross-linked dressings may have poor biological functions and ineffective tissue healing abilities, rendering them unsuitable for treating diabetic wounds. The encapsulation of bioactive compounds in these polymer-based dressings is a potential technique for tissue healing treatment, especially chronic wounds [56]. Antibiotics, growth factors, stem cells, plant extracts, antioxidants, anti-inflammatory medicines (e.g., curcumin, etc.), and vitamins are the examples of bioactive compounds which have been utilized in tissue healing applications. Wound dressings such as hydrogels, foams, membranes, films, nanofibers, transdermal patches, etc. are a few polymeric wound dressings that can hold bioactive compounds [57].

In a recent article by Ahmadian *et al.* [53], the authors notified the use and importance of synthetic polymers including PEG, PU, PVA, and PCL as well as natural polymers such as chitin, chitosan, and alginate. They further highlighted that the creation of innovative wound dressings is extremely important due to the drawbacks and shortcomings of gauzes and bandages as conventional wound dressings, including their inability to keep the wound moist, a requirement for frequent replacement, and painful separation [53]. The process of electrospinning is the one that is most frequently used to make wound dressings by using synthetic polymer [53, 58].

3. Conclusion

Electrospinning is one of the techniques to produce nanofibers that is widely explored for tissue healing application. There are several types of electrospinning variation to produce nanofiber membranes and scaffolds that require alteration in electrospinning parameters. A smooth projection of nanofibers that constructing 3D mat with interconnecting pores has capability to attract cells and tissue integration within the pores and between the nanofibers. More advanced and complex multi-fluid consist of natural and synthetic polymers electrospinning technology can be used in the future to create nanofibers with new shapes. Simultaneously, electrospun nanofibers can transport a range of active chemicals and can continually release medications, essential oils, plant extract, and nanoparticles, which are extremely advantageous for boosting the overall performance of tissue healing.

Acknowledgment

This study was supported by Hi-Tech F4 Program: Project III from Department of Deputy Vice Chancellor (Research & Innovation), Universiti Teknologi Malaysia, through the grant number Q.J130000.4609.00Q11.

Conflict of Interest

The authors declare no conflict of interest.

References

- [1] H. Lee and I. S. Kim, Nanofibers: Emerging progress on fabrication using mechanical force and recent applications, *Polymer Reviews*, 2018, 58(4): 688–716. <https://doi.org/10.1080/15583724.2018.1495650>.
- [2] J. Song, M. Kim and H. Lee, Recent advances on nanofiber fabrications: Unconventional state-of-the-art spinning techniques, *Polymers*, 2020, 12(6):1386. <https://www.mdpi.com/2073-4360/12/6/1386>.
- [3] D. Jao and V. Z. Beachley, Continuous dual-track fabrication of polymer micro-/nanofibers based on direct drawing, *ACS Macro Letters*, 2019, 8(5):588–595. <https://doi:10.1021/acsmacrolett.9b00167>.
- [4] S. Shin, F. Menk, Y. Kim, J. Lim, K. Char, R. Zental and T. L. Cho, Living light-induced crystallization-driven self-assembly for rapid preparation of semiconducting nanofibers, *Journal of the American Chemical Society*, 2018, 140(19):6088–6094. <https://doi:10.1021/jacs.8b01954>.
- [5] W. Qin, J. Li, J. Tu, H. Yang, Q. Chen and H. Liu, Fabrication of porous chitosan membranes composed of nanofibers by low temperature thermally induced phase separation, and their adsorption behavior for Cu²⁺, *Carbohydrate Polymer*, 2017, 178:338–346. <https://doi.org/10.1016/j.carbpol.2017.09.051>.
- [6] Z. Kamin, N. Abdulrahim, M. Misson, C. K. Chiam, R. Sarbatly, D. Krishnaiah and A. Bono, Use of melt blown polypropylene nanofiber templates to obtain homogenous pore channels in glycidyl methacrylate/ethyl dimethacrylate-based monoliths, *Chemical Engineering Communications*, 2021, 208(5):661–672. <https://doi.org/10.1080/00986445.2020.1715958>.
- [7] A. Luraghi, F. Peri and L. Moroni, Electrospinning for drug delivery applications: A review, *Journal of Controlled Release*, 2021, 334:463–484. <https://doi.org/10.1016/j.jconrel.2021.03.033>.
- [8] S. Sabra, D. M. Ragab, M. M. Agwa and S. Rohani, Recent advances in electrospun nanofibers for some biomedical applications, *European Journal of Pharmaceutical Sciences*, 2020, 144:105224. <https://doi.org/10.1016/j.ejps.2020.105224>.
- [9] S. Kumbar, R. James, S. Nukavarapu and C. Laurencin, Electrospun nanofiber scaffolds: Engineering soft tissues, *Biomedical Materials*, 2008, 3(3):034002. <https://doi.org/10.1088/1748-6041/3/3/034002>.
- [10] X. Liu, H. Xu, M. Zhang and D. G. Yu, Electrospun medicated nanofibers for wound healing, *Membranes (Basel)*, 2021, 11(10). <https://doi.org/10.3390/membranes11100770>.
- [11] A. Haider, S. Haider and I. -K. Kang, A comprehensive review summarizing the effect of electrospinning parameters and potential applications of nanofibers in biomedical and biotechnology, *Arabian Journal of Chemistry*, 2018, 11(8), 1165–1188. <https://doi.org/10.1016/j.arabjc.2015.11.015>.
- [12] H. R. Bakhsheshi-Rad, A. F. Ismail, M. Aziz, Z. Hadisi, M. Omid and X. Chen, Antibacterial activity and corrosion resistance of Ta₂O₅ thin film and electrospun PCL/MgO-Ag nanofiber coatings on biodegradable Mg alloy implants, *Ceramics International*, 2019, 45(9):11883–11892. <https://doi.org/10.1016/j.ceramint.2019.03.071>.
- [13] R. Nayak, R. Padhye, I. L. Kyrtziz, Y. B. Truong and L. Arnold, Recent advances in nanofibre fabrication techniques, *Textile Research Journal*, 2012, 82(2):129–147. <https://doi.org/10.1177/0040517511424524>.
- [14] S. Parham, A. Z. Kharazi, H. R. Bakhsheshi-Rad, H. Ghayour, A. F. Ismail, H. Nur and F. Berto, Electrospun nano-fibers for biomedical and tissue engineering applications, *Materials*, 2020, 13(9):2153. <https://www.mdpi.com/1996-1944/13/9/2153>.

- [15] K. Bootdee and M. Nithitanakul, Poly(d,l-lactide-co-glycolide) nanospheres within composite poly(vinyl alcohol)/aloe vera electrospun nanofiber as a novel wound dressing for controlled release of drug, *International Journal of Polymeric Materials and Polymeric Biomaterials*, 2021, 70(4):223–230. <https://doi.org/10.1080/00914037.2019.1706512>.
- [16] K. Schuhladden, S. N. V. Raghu, L. Liverani, Z. Neščáková and A. R. Boccaccini, Production of a novel poly(ϵ -caprolactone)-methylcellulose electrospun wound dressing by incorporating bioactive glass and Manuka honey, *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, 2021, 109(2):80–192. <https://doi.org/10.1002/jbm.b.34690>.
- [17] K. Ulubayram, S. Calamak, R. Shahbazi and I. Eroglu, Nanofibers based antibacterial drug design, delivery and applications, *Curr Pharm Des*, 2015, 21(15):1930–1943. <https://doi:10.2174/1381612821666150302151804>.
- [18] R. Naomi, H. Bahari, P. M. Ridzuan, and F. Othman, Natural-based biomaterial for skin wound healing (gelatin vs. collagen), *Polymers*, 2021, 13(14):2319. <https://www.mdpi.com/2073-4360/13/14/2319>.
- [19] J. Xia, H. Zhang, F. Yu, Y. Pei and X. Luo, Superclear, Porous cellulose membranes with chitosan-coated nanofibers for visualized cutaneous wound healing dressing, *ACS Applied Materials and Interfaces*, 2020, 12(21):24370–24379. <https://doi.org/10.1021/acsami.0c05604>.
- [20] D. N. Al-Jbour, D. M. Beg, J. Gimbut and M. A. K. M. Alam, An overview of chitosan nanofibers and their applications in the drug delivery process, *Current Drug Delivery*, 2019, 16(4):272–294. <https://doi.org/10.2174/1567201816666190123121425>.
- [21] R. Naomi, J. Ratanavaraporn and M. B. Fauzi, Comprehensive review of hybrid collagen and silk fibroin for cutaneous wound healing, *Materials*, 2020, 13(14):3097. <https://www.mdpi.com/1996-1944/13/14/3097>.
- [22] N. Chinatangkul, S. Tubtimsri, D. Panchapornpon, P. Akkaramongkolporn, C. Limmatvapirat and S. Limmatvapirat, Design and characterisation of electrospun shellac-polyvinylpyrrolidone blended micro/nanofibres loaded with monolaurin for application in wound healing, *International Journal of Pharmaceutics*, 2019, 562:258–270. <https://doi.org/10.1016/j.ijpharm.2019.03.048>.
- [23] M. Kurakula and G. S. N. K. Rao, Pharmaceutical assessment of polyvinylpyrrolidone (PVP): As excipient from conventional to controlled delivery systems with a spotlight on COVID-19 inhibition, *Journal of Drug Delivery Science and Technology*, 2020, 60:102046. <https://doi.org/10.1016/j.jddst.2020.102046>.
- [24] N. Raina, R. Pahwa, J. K. Khosla, P. N. Gupta and M. Gupta, Polycaprolactone-based materials in wound healing applications, *Polymer Bulletin*, 2021, 9. <https://doi:10.1007/s00289-021-03865-w>.
- [25] M. Buzgo, A. Mickova, M. Rampichova and M. Doupnik, Blend electrospinning, coaxial electrospinning, and emulsion electrospinning techniques, Core-shell Nanostructures for Drug Delivery and Theranostics, 2018, 325–347. <https://doi.org/10.1016/B978-0-08-102198-9.00011-9>.
- [26] J. Wu, Z. Zhang, J. Gu, W. Zhou, X. Liang, G. Zhou, C. C. Han, S. Xu and Y. Liu, Mechanism of a long-term controlled drug release system based on simple blended electrospun fibers, *Journal of Controlled Release*, 2020, 320:337–346. <https://doi.org/10.1016/j.jconrel.2020.01.020>.
- [27] M. M. Abdul Hameed, S. A. P. Mohamed Khan, B. M. Thamer, A. Al-Enizi, A. Aldalbahi, H. El-Hamshary and M. H. El-Newehy, Core-shell nanofibers from poly (vinyl alcohol) based biopolymers using emulsion electrospinning as drug delivery system for cephalexin drug, *Journal of Macromolecular Science, Part A*, 2020, 58(2):130–144. <https://doi.org/10.1080/10601325.2020.1832517>.
- [28] P. Coimbra, J. P. Freitas, T. Gonçalves, M. H. Gil and M. Figueiredo, Preparation of gentamicin sulfate eluting fiber mats by emulsion and by suspension electrospinning, *Materials Science and Engineering: C*, 2019, 94:86–93. <https://doi.org/10.1016/j.msec.2018.09.019>.
- [29] S. Su, T. Bedir, C. Kalkandelen, A. O. Başar, H. T. Şaşmazel, C. B. Ustundag, M. Sengor and O. Gunduz, Coaxial and emulsion electrospinning of extracted hyaluronic acid and keratin based nanofibers for wound healing applications, *European Polymer Journal*, 2021, 142:110158.

<https://doi.org/10.1016/j.eurpolymj.2020.110158>.

- [30] P. Rathore and J. D. Schiffman, Beyond the single-nozzle: Coaxial electrospinning enables innovative nanofiber chemistries, geometries, and applications, *ACS Applied Materials and Interfaces*, 2021, 13(1):48–66. <https://doi.org/10.1021/acsami.0c17706>.
- [31] P. Gupta and G. L. Wilkes, Some investigations on the fiber formation by utilizing a side-by-side bicomponent electrospinning approach, *Polymer*, 2003, 44(20):6353–6359. [https://doi.org/10.1016/S0032-3861\(03\)00616-5](https://doi.org/10.1016/S0032-3861(03)00616-5).
- [32] D. Li, M. Wang, W.-L. Song, D.-G. Yu and S. W. A. Bligh, Electrospun janus beads-on-A-string structures for different types of controlled release profiles of double drugs, *Biomolecules*, 2021, 11(5):635. <https://www.mdpi.com/2218-273X/11/5/635>.
- [33] X. Zhang, C. Chi, J. Chen, X. Zhang, M. Gong, X. Wang, J. Yan, R. Shi, L. Zhang and J. Xue, Electrospun quad-axial nanofibers for controlled and sustained drug delivery, *Materials and Design*, 2021, 206:109732. <https://doi.org/10.1016/j.matdes.2021.109732>.
- [34] J. Yin and L. Xu, Batch preparation of electrospun polycaprolactone/chitosan/aloe vera blended nanofiber membranes for novel wound dressing, *International Journal of Biological Macromolecules*, 2020, 160:352–363. <https://doi.org/10.1016/j.ijbiomac.2020.05.211>.
- [35] S. Fahimirad, H. Abtahi, P. Satei, E. Ghaznavi-Rad, M. Moslehi and A. Ganji, Wound healing performance of PCL/chitosan based electrospun nanofiber electrospayed with curcumin loaded chitosan nanoparticles, *Carbohydrate Polymers*, 2021, 259:117640. <https://doi.org/10.1016/j.carbpol.2021.117640>.
- [36] G. El Fawal, H. Hong, X. Mo and H. Wang, Fabrication of scaffold based on gelatin and polycaprolactone (PCL) for wound dressing application, *Journal of Drug Delivery Science and Technology*, 2021, 63:102501. <https://doi.org/10.1016/j.jddst.2021.102501>.
- [37] I. Unalan, S. J. Endlein, B. Slavik, A. Buettner, W. H. Goldmann, R. Detsch and A. R. Boccaccini, Evaluation of electrospun poly(ϵ -caprolactone)/gelatin nanofiber mats containing clove essential oil for antibacterial wound dressing, *Pharmaceutics*, 2019, 11(11). <https://doi.org/10.3390/pharmaceutics11110570>.
- [38] Y. Ghiyasi, E. Salahi and H. Esfahani, Synergy effect of *Urtica dioica* and ZnO NPs on microstructure, antibacterial activity and cytotoxicity of electrospun PCL scaffold for wound dressing application, *Materials Today Communications*, 2021, 26:102163. <https://doi.org/10.1016/j.mtcomm.2021.102163>.
- [39] C. L. Casper, J. S. Stephens, N. G. Tassi, D. B. Chase and J. F. Rabolt, Controlling surface morphology of electrospun polystyrene fibers: Effect of humidity and molecular weight in the electrospinning process, *Macromolecules*, 2004, 37(2):573–578. <https://doi.org/10.1021/ma0351975>.
- [40] A. P. Golin, Humidity effect on the structure of electrospun core-shell PCL-PEG fibers for tissue regeneration applications, Master thesis, Western University, 2014.
- [41] G. Liu, Z. Gu, Y. Hong, L. Cheng and C. Li, Electrospun starch nanofibers: Recent advances, challenges, and strategies for potential pharmaceutical applications, *Journal of Controlled Release*, 2017, 252:95–107. <https://doi.org/10.1016/j.jconrel.2017.03.016>.
- [42] S. Kailasa, M. S. B. Reddy, M. R. Maurya, B. G. Rani, K. V. Rao and K. K. Sadasivuni, Electrospun nanofibers: materials, synthesis parameters, and Their role in sensing applications, *Macromolecular Materials and Engineering*, 2021, 6(11):2100410. <https://doi.org/10.1002/mame.202100410>.
- [43] M. M. Demir, I. Yilgor, E. Yilgor and B. Erman, Electrospinning of polyurethane fibers, *Polymer*, 2002, 43(11):3303–3309. [https://doi.org/10.1016/S0032-3861\(02\)00136-2](https://doi.org/10.1016/S0032-3861(02)00136-2).
- [44] C. Mit-uppatham, M. Nithitanakul and P. Supaphol, Ultrafine electrospun polyamide-6 fibers: Effect of solution conditions on morphology and average fiber diameter, *Macromolecular Chemistry and Physics*, 2004, 205(17):2327–2338. <https://doi.org/10.1002/macp.200400225>.

- [45] M. A. Teixeira, M. T. P. Amorim and H. P. Felgueiras, Poly (vinyl alcohol)-based nanofibrous electrospun scaffolds for tissue engineering applications, *Polymers*, 2019, 12(1):7. <https://doi.org/10.3390/polym12010007>.
- [46] B. Ding, X. Wang and J. Yu, *Electrospinning: Nanofabrication and applications*, 1st ed., Netherlands: Elsevier, 2018.
- [47] S. Thakkar, *Introducing bioactivity into electrospun scaffolds for in situ cardiovascular tissue engineering*, PhD thesis, Technische Universiteit Eindhoven, 2018.
- [48] R. Thomas, K. R. Soumya, J. Mathew and E. K. Radhakrishnan, Electrospun polycaprolactone membrane incorporated with biosynthesized silver nanoparticles as effective wound dressing material, *Applied Biochemistry and Biotechnology*, 2015, 176(8):2213–2224. <https://doi.org/10.1007/s12010-015-1709-9>.
- [49] Z. Li and C. Wang, *Effects of working parameters on electrospinning*, in: *One-dimensional nanostructures*, Springer: Germany, 2013, 15–28.
- [50] S. Thomas, Y. Grohens and N. Ninan, *Nanotechnology applications for tissue engineering*, 1st ed., Netherlands: Elsevier, 2015.
- [51] A. Haider, S. Haider and I. -K. Kang, A comprehensive review summarizing the effect of electrospinning parameters and potential applications of nanofibers in biomedical and biotechnology, *Arabian Journal of Chemistry*, 2018, 11(8):1165–1188. <https://doi.org/10.1016/j.arabjc.2015.11.015>.
- [52] M. Sadia, M. A. M Zaki, S. K. Jaganathan, M. F. M. Shakhiih, A. S. Kamarozaman, N. Ablah and S. Saidin, Blending of *Moringa oleifera* into biodegradable polycaprolactone/silver electrospun membrane for hemocompatibility improvement, *Arabian Journal for Science and Engineering*, 2023, 4(6). <https://doi.org/10.1007/s13369-023-07736-6>.
- [53] Z. Ahmadian, H. Adiban, M. Rashidipour and M. R. Eskandari, Bioactive natural and synthetic polymers for wound repair, *Macromolecular Research*, 2022, 1–32. <https://doi.org/10.1007/s13233-022-0062-4>.
- [54] Z. Hussain, H. E. Thu, A. N. Shuid, H. Katas and F. Hussain, Recent advances in polymer-based wound dressings for the treatment of diabetic foot ulcer: An overview of state-of-the-art, *Current Drug Targets*, 2018, 19(5):527–550. <https://doi.org/10.2174/1389450118666170704132523>.
- [55] S. Alven and B. A. Aderibigbe, Fabrication of hybrid nanofibers from biopolymers and poly (vinyl alcohol)/poly (ϵ -caprolactone) for wound dressing applications, *Polymers (Basel)* 2021, 13(13). <https://doi.org/10.3390/polym13132104>.
- [56] X. Li, K. Nan, L. Li, Z. Zhang and H. Chen, In vivo evaluation of curcumin nanoformulation loaded methoxy poly(ethylene glycol)-graft-chitosan composite film for wound healing application, *Carbohydrate Polymers*, 2012, 88(1):84–90. <https://doi.org/10.1016/j.carbpol.2011.11.068>.
- [57] S. Alven, X. Nqoro and B. A. Aderibigbe, Polymer-based materials loaded with curcumin for wound healing Applications, *Polymers*, 2020, 12(10):2286. <https://www.mdpi.com/2073-4360/12/10/2286>.
- [58] P. Supaphol, O. Suwantong, P. Sangsanoh, S. Srinivasan, R. Jayakumar and S. V. Nair, Electrospinning of biocompatible polymers and their potentials in biomedical applications, *Biomedical Applications of Polymeric Nanofibers*, 2011, 213–239. https://doi.org/10.1007/12_2011_143.